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(54) Title: METHOD FOR MAKING WORT HAVING IMPROVED FILTERABILITY AND/OR INCREASED YIELD

(57) Abstract

The present invention provides worts with increased yield and improved filterability as well as a process for preparing said wort comprising the steps of: (a) preparing mash from malted or unmalted cereals, or a mixture of malted and unmalted cereals, in the presence of a mixture of enzyme activities, (b) filtering the mash so obtained to obtain the wort, wherein said mixture of enzyme activities is selected from a mixture comprising at least β -glucanase activity and α -L-arabinofuranose releasing activity or a mixture comprising at least β -glucanase activity and α -arabinofuranose releasing activity.

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Method for making wort having improved filterability and/or increased yield

Field of the invention

The present invention provides a process for making wort having improved filterability and/or showing increased brewing yield. The invention also provides for methods of brewing beer, making wine and potable alcohol from wort made according to the present invention. The invention also concerns the use of enzymes in improving filterability and/or yield of wort made from malted and/or unmalted cereals, as well as compositions comprising mixtures of enzyme activities according to the invention.

Background of the invention

The use of enzymes in making fermentable wort is known for a long time. For example, for brewing beer grains and/or malted grains are liquefied and saccharified in order to yield fermentable sugars. Liquefaction steps may be improved by the use of thermostable α-amylases as described for instance in US 4.285,975 or US 5,180,669. Also proteases are used to increase the amount of freely available nitrogen in the wort to improve fermentation. Alter filtration of the liquefied and saccharified mash, the obtained wort is inoculated with special strains of yeast (Saccharomyces sp.) which convert sugars into ethanol and characteristic flavour compounds.

However, apart from starch other polysaccharides are present in cereal grains. For example \(\beta\)-glucans are present (R.J. HENRY J.Sci Food Agric. (1985) 36, 1243-1253) as listed below for various cereals:

Cereal	% ß-glucans
wheat	0.5 - 8.0
barley	4.0 - 8.0
rye	5.0 - 10.0
rice	1.0 - 3.0
oat	5.0 - 10.0

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These \(\text{B-glucans} \) consist in \(\text{B-1,3/B-1,4} \) linkages between \(\text{B-D-glucopyranose} \) moieties (M.J.EDNEY, B.A. MARCHYLO and A.W. Mac GREGOR J. Inst. Brew. (1991) 97, 39-44). \(\text{B-glucans} \) are highly viscous (N.WAGNER \(et al. \) Monatschrift für Brauwissenschaft (1988) Heft 10, 384-395) and bring wort and beer filtration problems (S.AASTRUP Carlsberg Res. Commun. (1979) 44, 289-304) if they are not hydrolysed during the liquefaction step. This is the reason why

ß-glucanases from *Bacillus subtilis* (R.BORRISS Zeitschr. für allgem. Mikrobiol. (1981) 21(1), 7-17), *Penicillium emersonii* (A.P.MOLONEY, *et al.* Enzym. Microb. Technol. (1983) 5, 260-264), *Mucor miehei* (F.BRANISLAV European Patent 0 504 947 A2, 1992) or *Trichoderma* sp. (S.P. SHOEMAKER and R.D.BROWN Jr Biochim. Biophys. Acta (1978) 523, 133-146) are widely used at industrial scale, in addition to the β-glucanase present in the malt: the latter is not sufficiently thermostable to be active

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during the brewing diagramme processing (M.HRMOVA and G.B. FINCHER Biochem. J. (1993) 289, 453-461).

Non-starch polysaccharides also include pentosans, the structure of which have been widely studied recently (H.GRUPPEN et al. Carbohydr. Res. (1992) 233, 45-64; G.ANNISON et al., Carbohydr.

Polym. (1992) 19, 151-159; T. ITO et al., J. Carbohydr. Chem. (1994) 13(3) 491-498) in particular those of barley and malt (R.J. VIETOR et al., Carbohydr. Res. (1994) 254, 245-255;

R.J. VIETOR et al., Carbohydr. Polym. (1994) 24, 113-118). British patent 2,150,933 shows the interest for a pentosanase from *Penicillium emersonii* to improve the production and extraction of fermentable sugars in brewing. US Patent 4,746,517 demonstrates the high efficiency of the xylanolytic system from *Disporotrichum dimorphosporum* to improve wort and beer filterability.

The use of xylanase B to improve filterability of wort has also been mentioned in WO94/14965. This application is herein incorporated by reference.

Applications which may be mentioned in conjunction with the use of enzymes in making wort for fermentation are:

s WO94/21785.

Despite the advance which has been made in this area, there is still a need for methods of preparing wort with further improved filterability and/or higher yields, and enzyme preparations for use therein.

Summary of the invention

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The present invention provides worts with increased yield and improved filterability as well as a process for preparing said wort comprising the steps of:

- (a) preparing mash from malted or unmalted cereals, or a mixture of malted and unmalted cereals, in the presence of a mixture of enzyme activities,
- (b) filtering the mash so obtained to obtain the wort,

wherein said mixture of enzyme activities is selected from a mixture comprising at least β -glucanase activity and α -L-arabinofuranose releasing activity or a mixture comprising at least β -glucanase activity endo-xylanase activity and α -arabinofuranose releasing activity. Preferably, according to the process of the invention, arabinofuranose releasing activity is provided by an enzyme selected from α -L-arabinofuranosidase (EC 3.2.1.55) and (1->4)- β -D-arabinoxylan arabinofuranohydrolase (AXH), or a mixture of the said enzymes. The use of α -L-arabinofuranosidase or (1->4)- β -D-arabinoxylan arabinofuranohydrolase (AXH) obtainable from Aspergillus strain is still further preferred. According to one embodiment α -L-arabinofuranosidase A from Aspergillus niger is used.

According to another embodiment the invention provides a process for making beer, and/or potable alcohol, and other alcoholic beverages, wherein a wort is fermented obtainable according to the invention.

According to a further embodiment the invention envisages the use of α -L-arabinofuranosidase B from Aspergillus niger in a process of improving filterability of wort.

According to another embodiment the invention provides for the use of α -L-arabinofuranosidase A in a process of increasing yield of wort.

The invention also provides an enzyme preparation suitable for use in a process of making wort, said composition comprising a mixture of enzyme activities selected from a mixture comprising at least β -glucanase activity and α -arabinofuranose releasing activity or a mixture comprising at least β -glucanase activity endo-xylanase activity and α -arabinofuranose releasing activity.

Preferred according to the invention is an enzyme preparation, wherein said arabinofuranosyl releasing activity is provided for by an enzyme selected from α -L-arabinofuranosidase (EC 3.2.1.55) and (1->4)-B-D-arabinoxylan arabinofuranohydrolase (AXH), or a mixture thereof. Preferred according to the invention is an enzyme preparation wherein said α -L-arabinofuranosidase or said (1->4)-B-D-arabinoxylan arabinofuranohydrolase (AXH) is obtainable from Aspergillus strain.

Description of the invention

We have now surprisingly found that when wort is made in the presence of a mixture of enzyme activities comprising β-glucanase activity, α-L-arabinofuranosidase activity and preferably endo-β-1,4-xylanase, a liquefied and saccharified mash is obtained which can be filtered much easier than mash obtained using β-glucanases alone or mixtures of β-glucanases and endo-xylanase. Moreover, the wort obtained after filtration of the mash shows higher yield. Yield in this regard refers to the amount of fermentable sugars in the wort, expressed as a percentage of sugars present in the raw materials.

The yield improvement of the wort is dependent on the malt chosen. The malts exemplified herein show already good yields without the enzymes activities according to the invention. It is envisaged that more dramatic improvements may be obtained with poorer quality malts, with unmalted cereals or with mixed brews.

The manufacturing of the wort may be performed conventionally, involving liquefaction and saccharification of cereal material, usually with the aid of α -amylases and proteases, to obtain a liquefied and saccharified mash.

Suitable starting materials are cereals, either so-called raw or unmalted material or malted, or mixtures thereof (mixed brew). For example, cereals such as barley, wheat, corn, sorghum, oat and rice can be used, either malted or raw, or mixed. Preferably, the method according to the invention is used in 100% malt brew.

In the case of raw cereals, the liquefaction step usually comprises grinding of the cereal raw material to obtain a flour of suitable particle size, hydrating with from about 1 to about 4, preferably about 3 parts of water, and optionally, depending on the endoprotease used, from about 50 to about 300 ppm of calcium, preferably 200 ppm Ca²⁺. Enzymes from *Bacillus stearothermophilus* appear to be less Calcium-dependent. Consequently, no Ca²⁺ supplementation is required in that case. The particle size of the ground cereals should not exceed about 3 mm; not more than 3,5% should exceed 1,3 mm; not more than 1,5% should be smaller than 0.25 mm. Enzymes that may be used in addition are cellulases. B-glucanases, and or other plant cell wall degrading enzymes.

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The liquefaction medium is usually adjusted to a pH of between about 5 and 8, preferably between about 6 and 7, using, for example, calcium hydroxide. It is important to add α-amylase, preferably a thermostable α-amylase to the liquefaction medium as well as an endoprotease in a dosage sufficient to at least partially liquefy the cereal starch, and to at least partially degrade protein. Suitable dosages of α-amylase are from about 0,5 to about 2,0, preferably about 1 - 1,5 kg per Ton, when B.A.T.S. is used. Suitable dosages of proteases are, in the case of Brewers protease 2000, more than 0.5 kg/Ton grains (kg/T), preferably more than 1 kg/T. In the case of Panstimase 400 more than 2 kg/T, preferably more than 5, more preferably more than 10 kg/T should be used.

In the liquefaction process a number of steps are usually carried out at elevated temperature: after adding α-amylase and protease the mixture is maintained at a temperature between about 40°C and 65°C, preferably between about 45 and 55°C, most preferably 50°C, until a sufficient liquefaction is obtained. The time needed depends on the cereal or mixture of cereals used, but usually from about 30 minutes till about 2 hours is satisfactory. Subsequently, the temperature is raised gradually, the rate not being critical, till about 90-95°C and left at that temperature for about 30 minutes to about 1 hour. Then, the mixture is cooled to a temperature at which saccharification takes place: usually at about 50°C to about 70°C, preferably between about 55°C and 65°C, most preferably about 60°C. Slightly higher temperatures than 70°C should be possible, depending on the thermostability of the enzymes used in the saccharification step. When the preferred temperature is reached saccharifying enzymes are added, such as Brewers fermex (α-amylase) or Novamyl (recombinant β-amylase) in amounts usually ranging from about 400 g/T to about 1 kg/T for Brewers fermex. Also glucoamylases are frequently used. The saccharification takes from about 30 minutes to about 2 hours, whereafter the temperature is raised to about 75°C to about 85°C, *inter alia* to inactivate enzymes and unwanted microorganisms, and kept at the preferred elevated temperature for about 10 minutes; the period is not very critical.

The mash so obtained is subsequently filtered using equipment well known in the art; a funnel with Schleicher & Schuell paper filter works satisfactorily. After filtration, the wort is fermented by a suitable yeast, under conditions depending on the strain used, and the final purpose; in addition to brewing beer, production of alcohol as biofuel or as alcoholic beverage are envisaged by the instant invention. Suitable strains, and suitable conditions are well known to the person skilled in the art.

Other enzymes in the prepration of the wort that are conventionally used are \(\beta\)-glucanases. The \(\beta\)-glucanase is preferably thermostable enough to be active during the first step of a standard brewing diagramme (typically several minutes at 50°C and pH 5.6). Several microbial enzymes can fullfil this requirement e.g. \(\beta\)-glucanase from \(Penicillium\) emersonii, \(Trichoderma\) longibrachiatum, \(Bacillus\) amyloliquefaciens. Excellent results may be obtained with \(\beta\)-glucanase from \(Bacillus\) amyloliquefaciens commercially available from Gist-Brocades under the trademark Filtrase L (+), having an activity of 600 BGR/g. Preferably, \(\beta\)-glucanase is obtained from a recombinant strain of \(Bacillus\) amyloliquefaciens. Usually also proteases may be used, such as Brewer's Protease and the like.

Endo-xylanases that may be used have been described in the section background art. The patent specifications mentioned there are herein incorporated by reference.

Endo-β-1,4-xylanase (preferably isoenzyme endo I (R.A. FOURNIER Biotechnology and Bioeng.(1985) XXVII, 539-546) and α-L-arabino-furanosidase (preferably isoenzyme A (F.J.M. KORMELINK *et al.*, Carborhydr. Res. (1993) 24, 345-353)) may be obtained from wild-type, mutated or recombinant strains of *Aspergillus niger*.

As regards the α -L-arabinose releasing activities, several enzymes may be envisaged. We have shown that AXH leads to better filterability. Two α -L-arabinosidases from Aspergillus niger appeared to lead to significantly higher filtration rates. As the mode of action of AXH, α -L-arabinosidase A and B all differ, it may be expected, that a mixture of the three arabinose releasing activities may yield to still better filtration rates.

Using the process according to the invention substantially improved filtration rates may be obtained of the liquefied and saccharified mash. This brings advantages in terms of a speedier process, less clogging of filters, and larger wort volumes.

Also the improved yield leads to a more economic brewing process. The wort according to the invention can be used in brewing beer, or making potable alcohol or biofuel, or in a process of making other alcoholic beverages. The practicing of the invention and the associated advantages are illustrated in greater detail in the following non-limitative Examples.

Experimental part

1/B-glucanase

In the Examples \(\mathbb{B}\)-glucanase was used from \(\textit{Bacillus amyloliquefaciens} \) commercially available from \(\text{Gist-Brocades under the trademark Filtrase L (+), having an activity of 600 BGR/g. \)

2/Endo-ß-1,4-xylanase

Endoxylanase was obtained from a pure culture of Aspergillus niger in a sterile tank and medium. The culture medium contains appropriate carbon and nitrogen sources just as mineral salts. The fermentation is carried out at a constant temperature between 30-40°C and pH is maintained within the range 3-5. The activity of the enzyme is measured by the hydrolysis of xylan from oat spelts suspended (35g/l) in 1M glycine buffer pH 2,75. the viscosity of this solution is determined by using a capillary viscometer (Ubbelhode type) at 47°C. The time dt needed for the upper menisk of the liquid to fall down between two reference points is measured within time T. The slope of the plot T versus 1/dt yields an apparent kinetic constant. 1 Lyx unit is the amount of enzyme needed to reach a value of 1 min-1 for that kinetic constant.

3/a-L-arabinofuranosidase

Isoenzyme A or isoenzyme B (F.J.M. KORMELINK et al., Carborhydr. Res. (1993) 24, 345-353) or Arabinoxylanhydrolase (F.J.M. KORMELINK et al., Appl. Microbiol. Biotechnol. (1991) 35, 753-758) have been obtained from a culture of recombinant Aspergillus niger or Aspergillus nidulans strains. The amino acid sequences, their encoding DNA, as well as the recombinant production of arabinofuranosidase A and B is described in detail in European patent application

0 506 190 A1, published on September 30, 1992, particularly in the Examples and Figures and the Sequence Listing, which relevant parts are herein incorporated by reference.

Activity of isoenzymes A and B is measured by the hydrolysis of p-nitrophenyl-α-L-arabinofuranoside. 1 ARF unit is the amount of enzyme needed to liberate 1 μmole p-nitrophenol per minute under the conditions of the test described in (Z. GUNATA et al., J. Agric. Food Chem. (1989) 38, 772).

Example 1

Wort was prepared from malt ground according to standard specifications for lauter tun filtration. Malt was ground with the EBC MIAG mill according to standard specifications, i.e. yielding a difference in fine (1 mm) to coarse (2.5 mm) extract in the range 1.5-2 %. One part malt is hydrated with 3 parts water or aqueous solution of enzyme at 50°C. This temperature is maintained during 20 minutes; it is then raised up to 63°C (1°C/min) and maintained at that temperature during 30 minutes. The medium is then heated at 72°C (1°C/min) and maintained at this temperature during 20 minutes. It is finally heated at 76°C and maintained at that temperature for 5 minutes. Water is added to compensate for water evaporation.

The mash is poured into a funnel containing Schleicher and Schuell paper filter. The volume of filtered wort is measured after 15 minutes. Specific gravity is determined at the end of the filtration. This value allows to calculate extract and yield.

1st serie (British malt).

5 brews have been carried out with different enzymes as shown in Table 1:

	Brew n°	Enzymatic u	inits/kg malt			
		BGR	LYX	ARF-A	AXH	ARF-B
25						
	1	0	0 .	0	0	0
	2	90	1200	0	0	0
	3	90	1200	1500	0	0
	4	90	1200	•	0	1500
30	5	90	1200	0	1500	0

In brew n°3, ARF is from α-L-arabinofuranosidase isoenzyme A

In brew n°4, ARF is from α-L-arabinofuranosidase isoenzyme B

In brew n°5, the arabinoxylanhydrolase AXH (F.J.M. KORMELINK et al. Appl. Microbiol. Biotechnol. (1991) 35, 753-758) is used to hydrolyse the arabinose moieties of arabinoxylans. The dose was 0,15 mg pure AXH per kg malt (AXH is not active on p-nitrophenyl-α-L-arabinofuranoside, therefore AXH has no ARF activity).

Results are presented in Table 2:

	Brew nº	Volume filtered after 15 minutes (ml)	Yield (%)	
5	1	62	83,04	
	2	88	83,69	
	3	102	83,64	
	4	106	83,24	
10	5	100	83,44	

2nd serie (French malt)

The same brews as described in the 1st series have been produced from a french malt. Results are presented in Table 3:

20	Brew n°	Volume filtered after 15 minutes (ml)	Yield (%)	,
	1	54	82,35	
	2	82	82,50	
	3	88	83,71	
25	4	92	82,60	
	5	88	82,50	

From these 2 series, the combination β -glucanase + endoxylanase + α -L-arabinofuranosidase isoenzyme B performs best to improve wort filtration, whereas the same combination in which α -L-arabino-furanosidase isoenzyme A replaces α -L-arabinofuranosidase isoenzyme B performs best to improve the yield, the filtration being also highly improved in comparison with the blank (brew 1).

Example 2

Worts were produced the same way as in example 1, always with the same batch of malt but varying the composition of the enzymes' mixture. 12 brews were carried out according to an experimental design in order to determine the role of each component of the mixture with regards to yield and filtration improvement. In all trials, α-L-arabinofuranosidase was isoenzyme A from A. niger.

All runs performed are presented in Table 4:

Brew no	e° Enzymatic units/kg malt			
	BGR	LYX	ARF (isoenzyme A	.)
1	30	200	200	
•		300	300	
2	90	300	300	
3	30	1200	300	
4	90	1200	300	
5	30	300	1500	
6	90	300	1500	
7	30	1200	1500	
8	90	1200	1500	
9	60	750	900	
10	60	750	900	
11	60	750	900	
12	0	0	0	•

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Results are shown in Table 5:

Brew n°	Volume filtered	Yield
	after 15 minutes (ml)	(%)
1	115	80,16
2	100	80,21
3	116	10,08
4	110	80,81
5	116	80,11
6	128	80,26
7	122	80,01
8	142	80,56
9	116	80,06
10	120	80,11
11	117	80,11
12	89	80,01

From Brews n°9-10-11 standard deviations may be determined:

2,1 ml for volume filtered after 15 minutes 0,03% for the yield

From Brews n°1 to 8, the effect of each component just as those of interactions between components may be determined:

	Cause	Effe	ect on
		Filterability	Yield
5	BGR	+2,75	+0,39
	LYX	+7,75	+0,16
	ARF	+16,75	-0,06
	BGR*LYX	+4,25	+0,29
	BGR*ARF	+13,25	-0,04
10	LYX*ARF	+2,75	-0,06

Only figures in bold type characters may be considered as significant (> 95%). α -L-arabinofuranosidase A alone and in combination with β -glucanase brings the strongest effect in filtration improvement, whereas β -glucanase, endoxylanase alone and in combinations contribute significantly to the yield improvement.

Example 3

In that serie, the same combination of enzymes is used:

60 BGR/kg malt

- + 750 LYX/kg malt
- + 900 ARF/kg malt

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but different malt were brewed with and without this enzyme combination. Worts were produced according to the same procedure as in Example 1.

Results are presented in Table 5:

Malt's	Volum	e filtered	Yie	ld
origin	after 1	5 minutes	(%	6)
	(m	1)		
	without	with	without	with
	enzyme	enzyme	enzyme	enzyme
				
USA (1)	131	171	79,75	79,77
USA (2)	114	164	83,74	83,97
UK	117	167	80,18	83,21
Canada (1)	106	153	81,42	81,52
Canada (2)	99	137	81,30	81,86
France	89	118	80,18	80,09

Thus, the mixture of enzyme activities is efficient for filtration improvement, whatever the malt brewed. The effect on yield is low but this is mainly because the malts used are already of a very high quality.

Example 4

Wheat is used as crude adjunct in several breweries. This cereal contains a high amount of pentosans. This is one reason to test the above mentioned combination of enzymes in a mixed brew.

Worts were produced according to the same procedure as in example 1 but 20% malt was replaced by 20% wheat.

Table 6 describes all runs performed:

Brew n°	Enzymatic units/kg malt				
	BGR	LYX	ARF		
1	0	0	. 0		
2	90	0	0		
3	0	120	0		
4	0	120	300		
5	0	120	1500		
6	90	120	1500		

Results are given in Table 7:

o	Brew n°	Volume filtered after 15 minutes (ml)	Yield (%)	
	1	66	81,01	
	2	89	81,52	
	3	62	80,81	
5	4	69	80,91	
	5	80	80,91	
	6	99	81,52	

These results confirm the interest for the combination described in the invention in the case of brews involving wheat adjuncts.

Example 5

In this example, wort was prepared from crude barley grains, variety PLAISANT.

Barley grains were ground with the EBC MIAG mill in order to make filter press type barley flour. 57g barley flour are added in 300ml water or aqueous solution of enzymes at 50°C. This temperature is maintained for 1h; it is then heated up to 63°C (1°C/min) and maintained at that temperature during 30 minutes. The medium is then heated up to 90°C (1°C/min) and maintained at that temperature during 20

10

15

minutes. Water is added to compensate for water evaporation. The mash is then poured into a funnel containing Schleicher and Schuell paper filter.

6 brews have been carried out with different enzymes as shown in Table 8 (ARF when mentioned is from Ara A):

Brew	Enzym	es	
No.	Microbial origin	Units/kg barley	
ı	-	0	
2(*)	Bacillus amyloliquefaciens + Aspergillus niger	90 BGR + 1200 Lyx + 1500 ARF	
3(*)	idem	90 BGR + 12000 Lyx + 15000 ARF	
4	Penicillium emersonii (**)	36 XVU	
5	Bacillus amyloliquefaciens + Disporotrichum dimorphosporum (***)	90 BGR + 120 XVU	
6	Trichoderma viride (****)	36 XVU	

- (*) according to the invention
- (**) commercially available from Gist-brocades under the trade name Filtrase® NL
- (***) commercially available from Gist-brocades under the trade name Filtrase® Br
- (****) commercially available from Gist-brocades under the trade name Natugrain®

Results are presented in Table 9.

Brew No.	Volume (ml) filtered after 15 minutes
1	111
2	117
3	164
4	141
5	172
6	136

These results show that the combination described in the invention is at least as good or even better performing than existing preparations in the case of brews involving 100% unmalted barley.

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Claims

- 1. A process for preparing wort comprising the steps of:
- (a) preparing mash from malted or unmalted cereals, or a mixture of malted and unmalted cereals, in the presence of a mixture of enzyme activities,
- (b) filtering the mash to obtain the wort,

wherein said mixture of enzyme activities is selected from a mixture comprising at least β -glucanase activity and α -arabinofuranose releasing activity or a mixture comprising at least β -glucanase activity endo-xylanase activity and α -arabinofuranose releasing activity.

- 2. A process according to claim 1, wherein said arabinofuranose releasing activity is provided by an enzyme selected from α-L-arabinofuranosidase (EC 3.2.1.55) and (1->4)-β-D-arabinoxylan arabinofuranohydrolase (AXH), or a mixture of the said enzymes.
- 3. A process according to claim 1, wherein said α -L-arabinofuranosidase or said (1->4)- β -D-arabinoxylan arabinofuranohydrolase (AXH) is obtainable from an Aspergillus strain.
 - 4. A process according to claim 3, wherein said α -L-arabinofuranosidase is α -L-arabinofuranosidase A from Aspergillus niger.
 - 5. A process according to claim 3, wherein said α -L-arabinofuranosidase is α -L-arabinofuranosidase B from Aspergillus niger.
- 6. A process for making beer and/or potable alcohol, wherein a wort is fermented obtainable according to any one of claims 1 to 5.
 - 7. Use of α -L-arabinofuranosidase B from Aspergillus niger in a process of improving filterability of wort.
- 30 8. Use of α-L-arabinofuranosidase A in a process of increasing yield of wort.
 - 9. An enzyme preparation suitable for use in a process of making wort, said composition comprising a mixture of enzyme activities selected from a mixture comprising at least β -glucanase activity and α -arabinofuranose releasing activity or a mixture comprising at least β -glucanase activity endo-xylanase activity and α -arabinofuranose releasing activity.
 - 10. An enzyme preparation according to claim 9, wherein said arabinofuranosyl releasing activity is provided for by an enzyme selected from α -L-arabinofuranosidase (EC 3.2.1.55) and (1->4)- β -D-arabinoxylan arabinofuranohydrolase (AXH), or a mixture thereof.

11. An enzyme preparation according to claim 10, wherein said α -L-arabinofuranosidase or said (1->4)- β -D-arabinoxylan arabinofuranohydrolase (AXH) is obtainable from an Aspergillus strain.

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A. CLASS IPC 6	SIFICATION OF SUBJECT MATTER C12C7/04 C12C5/02 C12P7	/06	
According	to International Patent Classification (IPC) or to both national	classification and IPC	
B. FIELD	S SEARCHED		
IPC 6	documentation searched (classification system followed by class C12C C12P	ification symbols)	
Documenta	ation searched other than minimum documentation to the extent	that such documents are included in	n the fields searched
Electronic	data base consulted during the international search (name of dat	a base and, where practical, search	terms used)
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X Furt	ther documents are listed in the continuation of box C,	X Patent family member	s are listed in annex,
'A' docum	tegories of cited documents: ent defining the general state of the art which is not lered to be of particular relevance	or priority date and not in cited to understand the pri	after the international filing date a conflict with the application but inciple or theory underlying the
"E" carlier filling o "L" docume	document but published on or after the international	involve an inventive step	el or cannot be considered to when the document is taken alone
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	actual completion of the international search	'&' document member of the i	
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Name and n	nailing address of the ISA European Patent Office, P.B. 5818 Patentiaan 2 NL - 2280 HV Rijswijk	Authorized officer	
	Tel. (+31-70) 340-2040, Tx. 31 651 epo nl. Fax: (+31-70) 340-3016	Bevan, S	

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